## **Research Note**

## Seroprevalence and risk factors for *Toxoplasma gondii* in sheep and goats in Jinzhou, Northeastern China

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Abstract. In the present study, serum samples from 402 sheep and 216 goats were collected from 5 counties in Jinzhou from August to October 2012 and antibodies to *Toxoplasma gondii* were detected by modified agglutination test (MAT). Overall, 104 (16.8%) had antibodies to *T. gondii* with antibody titres of 1:25 to 1:800. Seropositive samples were distributed in all the 5 counties and seroprevalences of *T. gondii* varied significantly with flock size, age and rearing system, but not with breed, gender and farm location. The seroprevalences in small farms (18.3%, 95/518, 95% confidence interval [CI], 15.0-21.7%) were statistically higher than that in large farms (9%, 9/100, 95% CI, 3.4-14.6%) (P < 0.05), older animals were statistically higher than that in younger animals (P < 0.01). The prevalence in extensively raised animals (P < 0.01). Small flock size and extensive rearing system are the potential risk factors for the prevalence of *Toxoplasma* infection in sheep and goats in Jinzhou. This is the first report of *T. gondii* and the risk factors.

Toxoplasma gondii is a protozoan parasite that infects up to a third of the world's population. Infection is mainly acquired by ingestion of food or water that is contaminated with oocysts shed by cats or by eating undercooked or raw meat containing tissue cysts (Dubey & Dubey, 2010; Hide et al., 2009; Montova & Liesenfeld, 2004). Infection of sheep and goats with T. gondii may cause early embryonic death, fetal death and mummification, abortion, stillbirth and neonatal death and thus can be responsible for heavy economic losses (Dubey & Dubey, 2010). The ingestion of undercooked lamb containing tissue cysts of T. gondii is considered a significant source

of toxoplasmosis in humans (Cook *et al.*, 2000; Dubey, 2000; Gao *et al.*, 2012; Jones *et al.*, 2009). People in Jinzhou have the habit of eating under-cooked 'barbecue', 'kabob' and 'instantly boiled mutton', making increased risk of toxoplasmosis.

Jinzhou is the central city of west part of Liaoning Province, northeastern China, which covers an area of 10,301 square kilometres with a population of 3.07 million. Jinzhou is lying to the north of China's Bohai Sea and the south of Mount Yiwuly, which has a humid continental climate and a relatively large variation in temperature over the course of a year; there are 4 distinct seasons and an annual average rainfall ranging from 540 to 640 mm, heavily concentrated in July and August alone. The geographic and natural climatic conditions are suitable for the development of agriculture, forestry, and livestock production. The livestock industry is an important economic resource in Jinzhou, which has approximately 420,000 sheep and goats. None of the animals had ever been vaccinated against toxoplasmosis; indeed, such vaccination is not practiced in China.

The present study was conducted to investigate the prevalence of *T. gondii* infection and to further explore the potential risk posed to humans in northeastern China.

Serum samples were collected from 618 sheep and goats via a jugular vein in Jinzhou (40°49'–42°08'N, 120°42'–122°36'E), including Heishan, Beizhen, Linghai, Yixian and Taihe from August to October 2012. Whenever possible, data regarding breed, age, gender, location, flock size, rearing system (extensive: daily grazing in favourable weather conditions and returning to fold at night or daily grazing with possibility of shelter in bad weather; intensive: sheep housed day and night) of each animal was collected (Table 1). Blood samples were centrifuged (3,000 rpm) for 5 min and stored at -20°C until use.

The modified agglutination test (MAT) has been evaluated extensively in experimentally and naturally infected sheep and goats, and is demonstrated sensitive and specific for assaying T. gondii antibodies in animals (Alvarado-Esquivel, Estrada-Malacon et al., 2013; Chikweto et al., 2011; Ragozo et al., 2008). In the present study, sera were diluted to 1:25, tested for T. gondii antibodies (IgG) by MAT as described previously (Dubey & Dubey, 2010). In brief, the harvested parasites were kept in 6% formaldehyde solution at 4°C overnight, and suspended in the alkaline buffer at 20,000 parasites/ml. Two-fold dilutions of sera were performed using the serum diluting buffer, starting with 1:25. Agglutination was done in V-bottom 96-well microtiter plates using a mixture of 50 µl antigen and 50 µl diluted sera. The plates were incubated at 37°C overnight. The test was considered positive when a layer of agglutinated parasites was formed

in wells at dilutions of 1:25 or higher; positive and negative controls were included in each test.

Differences in seroprevalence of infected sheep and goats and among associated factors were analyzed using Fisher's exact test in SAS statistical software (Version 9.3; SAS Institute Inc., Cary, NC, USA), 95% confidence intervals (CI) are given. Differences between levels within factors and interactions were considered to be statistically significant and highly significant when P < 0.05 and P < 0.01, respectively.

Antibodies to *T. gondii* were found in 72 (17.9%) of 402 sheep and 32 (14.8%) of 216 goats in titers of 35 sera with a titer of 25, 24 of 50, 17 of 100, 12 of 200, 9 of 400, and 7 of 800 or higher.

The results of the univariate analysis are shown in Table 1. 16.8% of the 618 tested sheep and goats were seropositive for *T. gondii* by MAT, which was higher than that reported by others (Wang CR et al., 2011; Xu et al., 2014; Zhao et al., 2011). The difference could be associated with management of the sampled animals, ecological conditions, life styles of inhabitants, climates, serological technique used, husbandry practice and the numbers of cats and rodents present. In addition, the feeding conditions and animal welfare are also the risk factors for *T. gondii* infection in sheep and goats (Dubey & Dubey, 2010; Wang CR et al., 2011).

The logistic regression showed that all the factors (breed, sex, age, location, flock size and rearing system) reported in the present study affected prevalence of infection (Table 1). The seroprevalence in sheep (17.9%, 72/402, 95% CI, 14.2-21.7%) was higher than that in goats (14.8%, 32/216, 95% CI, 10.1-19.6%), but the difference was not statistically significant (P > 0.05). Although the prevalence of T. gondii infection in sheep and goats across the world is variable, the literature generally indicates that the prevalence of infection amongst sheep is higher than that in goats (Kamani, Mani & Egwu, 2010; Lopes et al., 2013). The seroprevalence in female animals (18.3%, 81/443, 95% CI, 14.7-21.9%) was not found to be

Characteristics	No. examined	No. positive	Prevalence, % (95% CI)	Р
Breed				0.323
Sheep	402	72	17.9 (14.2,21.7)	
Goat	216	32	14.8 (10.1,19.6)	
Age(year)				0.001
yr≤1	86	4	4.7(1.3,11.4)	
1 <yr<u>&lt;3</yr<u>	307	50	16.3 (12.2,20.4)	
3 <yr< td=""><td>225</td><td>50</td><td>22.2(16.8,27.7)</td><td></td></yr<>	225	50	22.2(16.8,27.7)	
Gender				0.124
Male	175	23	13.1 (8.1,18.2)	
Female	443	81	18.3 (14.7,21.9)	
Location (County)				0.591
Heishan	156	31	19.9 (13.6,26.1)	
Beizhen	125	23	18.4 (11.6,25.2)	
Yixian	132	22	16.7 (10.3,23.0)	
Linghai	108	14	13.0 (6.7,19.3)	
Taihe	97	14	14.4 (7.4,21.4)	
Flock size				0.022
$Large(n \ge 100)$	100	9	9.0(3.4,14.6)	
Small(n<100)	518	95	18.3(15.0,21.7)	
Rearing system				0.005
Extensive	58	15	25.9(14.6, 37.1)	
$SIR^{b}$	400	74	18.5(14.7,22.3)	
IR <sup>c</sup>	160	15	9.4(4.9,13.9)	
Total	618	104	16.8 (13.9,19.8)	

Table 1. General characteristics of the 618 sheep and goats studied and seroprevalence of T. gondii infection<sup>a</sup> in Jinzhou, northeastern China

<sup>a</sup>The difference was considered significant when P value less than 0.05; CI, Confidence interval; <sup>b</sup>SIR, semiintensively raised, <sup>c</sup>IR, intensively raised.

significantly higher (P > 0.05) than that in males (13.1%, 23/175, 95% CI, 8.1–18.2%). This gender-related tendency of prevalence had been reported previously (Wang CR *et al.*, 2011; Xu *et al.*, 2014), however, a study in Nigeria (Kamani *et al.*, 2010) did not find an association between seroprevalences and genders.

In the present study, seroprevalence in sheep and goats increased progressively with age, ranging from 4.7%–22.2%, with the highest of 22.2% in samples which were >3 year old, the seroprevalences were statistically significantly higher than that

in younger groups ( $\leq$ 1-year-old) (P < 0.01). These results are similar to those of previous investigations (Alvarado-Esquivel, Silva-Aguilar, Villena, & Dubey, 2013; Ramzan *et al.*, 2009), suggesting the possibility of horizontal transmission in the investigated herds.

In the present survey, the seroprevalence in small farms (18.3%, 95/518, 95% CI, 15.0– 21.7%) was statistically higher (P < 0.05) than that in large farms (9%, 9/100, 95% CI, 3.4– 14.6%). In Jinzhou, sheep and goats feeding is predominated by small-scale rearing by farmer households, compared with other district. Thus, sheep have more chance to ingest the oocysts of T. gondii excreted by infected cats in poor breeding conditions of small farms. In addition, sheep and goats are raised extensively in small farms or semiintensively by individual families in the study region. And the prevalence in intensively raised sheep and goats was statistically lower (P < 0.01) than that in extensively and semi-intensively raised samples (Table 1). Our findings are similar to those of previous reported (Ragozo et al., 2008; Wang CR et al., 2011). The main reason for such a difference may be that, compared with extensively or semi-intensively raised animals, intensively raised sheep and goats are caged and thus have less chance to ingest the oocysts of T. gondii excreted by infected cats.

The prevalence of T. gondii infection varied from 14.4% to 19.9% among different regions in Jinzhou district, and the differences among seroprevence of T. gondii in different regions are shown in Table 1. Higher prevalence was found in Heishan (19.9%, 31/156, 95% CI, 13.6–26.1%), compared with other regions. Previous studies have shown that T. gondii infection in free-range chickens was a good indicator of the environmental contamination with oocysts because chickens became infected mainly by feeding from ground, feed, or soil contaminated with oocysts (Beltrame et al., 2012; Dubey, Lenhart, et al., 2005; Dubey, Rajapakse, Ekanayake, Sreekumar & Lehmann, 2005). Although the data collected in our study did not include the positive association between the presence of freerange chickens and T. gondii infection in sheep, previous study had shown that a high rate (20%) of T. gondii infection was found in free-range chickens from Heishan (Xu et al., 2012), which may contribute to the higher prevalence of T. gondii infection in sheep and goats from the same region.

Based on the results obtained in this study, it can be concluded that small flock size and extensive rearing system are the potential risk factors for *Toxoplasma* infection in sheep and goats in Jinzhou, northeastern China. Control and prophylactic measures should be adopted to improve the rearing system and the implementation of health promoting programs in a joint effort between farmers, farmers' associations and veterinarians to inform about the means of transmission of the infection and for a better understanding of toxoplasmosis.

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## REFERENCES

- Alvarado-Esquivel, C., Estrada-Malacon, M.A., Reyes-Hernandez, S.O., Perez-Ramirez, J.A., Trujillo-Lopez, J.I., Villena, I. & Dubey, J.P. (2013). Seroprevalence of *Toxoplasma gondii* in domestic sheep in Oaxaca State, Mexico. *Journal of Parasitology* **99**(1): 151-152.
- Alvarado-Esquivel, C., Silva-Aguilar, D., Villena, I. & Dubey, J.P. (2013). Seroprevalence of *Toxoplasma gondii* infection in dairy goats in Michoacan State, Mexico. *Journal of Parasitology* **99**(3): 540-542.
- Beltrame, M.A., Pena, H.F., Ton, N.C., Lino, A.J., Gennari, S.M., Dubey, J.P. & Pereira, F.E. (2012). Seroprevalence and isolation of *Toxoplasma gondii* from free-range chickens from Espirito Santo state, southeastern Brazil. *Veterinary Parasitology* 188(3-4): 225-230.
- Chikweto, A., Kumthekar, S., Tiwari, K., Nyack, B., Deokar, M.S., Stratton, G. & Dubey, J.P. (2011). Seroprevalence of *Toxoplasma gondii* in pigs, sheep, goats, and cattle from Grenada and Carriacou, West Indies. *Journal of Parasitology* **97**(5): 950-951.

- Cook, A.J., Gilbert, R.E., Buffolano, W., Zufferey, J., Petersen, E., Jenum, P.A. & Dunn, D.T. (2000). Sources of toxoplasma infection in pregnant women: European multicentre case-control study. European Research Network on Congenital Toxoplasmosis. *British Medical Journal* **321**(7254): 142-147.
- Dubey, J.P. (2000). Sources of *Toxoplasma* gondii infection in pregnancy. Until rates of congenital toxoplasmosis fall, control measures are essential. *British Medical* Journal **321**(7254): 127-128.
- Dubey, J.P. (2010). Toxoplasmosis of animals and humans (2nd ed.). Boca Raton: CRC Press. p.17-231.
- Dubey, J.P., Lenhart, A., Castillo, C.E., Alvarez, L., Marcet, P., Sreekumar, C. & Lehmann, T. (2005). *Toxoplasma gondii* infections in chickens from Venezuela: isolation, tissue distribution, and molecular characterization. *Journal of Parasitology* **91**(6): 1332-1334.
- Dubey, J.P., Rajapakse, R.P., Ekanayake, D.K., Sreekumar, C. & Lehmann, T. (2005). Isolation and molecular characterization of *Toxoplasma gondii* from chickens from Sri Lanka. *Journal of Parasitology* **91**(6): 1480-1482.
- Gao, X.J., Zhao, Z.J., He, Z.H., Wang, T., Yang, T.B., Chen, X.G. & Lun, Z.R. (2012). *Toxoplasma gondii* infection in pregnant women in China. *Parasitology* **139**(2): 139-147.
- Hide, G., Morley, E.K., Hughes, J.M., Gerwash, O., Elmahaishi, M.S., Elmahaishi, K.H. & Smith, J.E. (2009). Evidence for high levels of vertical transmission in *Toxoplasma gondii*. *Parasitology* **136**(14): 1877-1885.
- Jones, J.L., Dargelas, V., Roberts, J., Press, C., Remington, J.S. & Montoya, J.G. (2009). Risk factors for *Toxoplasma gondii* infection in the United States. Clinical infectious diseases: an official publication of the Infectious Diseases Society of America **49**(6): 878-884.

- Kamani, J., Mani, A.U. & Egwu, G.O. (2010). Seroprevalence of *Toxoplasma gondii* infection in domestic sheep and goats in Borno state, Nigeria. *Tropical Animal Health and Production* 42(4): 793-797.
- Lopes, A.P., Dubey, J.P., Neto, F., Rodrigues, A., Martins, T., Rodrigues, M. & Cardoso, L. (2013). Seroprevalence of *Toxoplasma* gondii infection in cattle, sheep, goats and pigs from the North of Portugal for human consumption. *Veterinary Para*sitology **193**(1-3): 266-269.
- Montoya, J.G. & Liesenfeld, O. (2004). Toxoplasmosis. *Lancet* **363**(9425): 1965-1976.
- Ragozo, A.M., Yai, R.L., Oliveira, L.N., Dias, R.A., Dubey, J.P. & Gennari, S.M. (2008).
  Seroprevalence and isolation of *Toxoplasma gondii* from sheep from Sao Paulo state, Brazil. *Journal of parasitology* **94**(6): 1259-1263.
- Ramzan, M., Akhtar, M., Muhammad, F., Hussain, I., Hiszczynska-Sawicka, E., Haq, A.U. & Hafeez, M.A. (2009). Seroprevalence of *Toxoplasma gondii* in sheep and goats in Rahim Yar Khan (Punjab), Pakistan. *Tropical Animal Health and Production* **41**(7): 1225-1229.
- Wang, C.R., Qiu, J.H., Gao, J.F., Liu, L.M., Wang, C., Liu, Q. & Zhu, X.Q. (2011). Seroprevalence of *Toxoplasma gondii* infection in sheep and goats in northeastern China. *Small Ruminant Research* 97: 130-133.
- Xu, P., Li, X., Guo, L., Li, B., Wang, J., Yu, D. & Liu, X.G. (2014). Seroprevalence of *Toxoplasma gondii* infection in Liaoning cashmere goat from northeastern China. *Parasite* 21: 22.
- Xu, P., Song, X., Wang, W., Wang, F., Cao, L. & Liu, Q. (2012). Seroprevalence of *Toxo*plasma gondii infection in chickens in Jinzhou, northeastern China. Journal of Parasitology **98**(6): 1300-1301.
- Zhao, G.H., Zhang, M.T., Lei, L.H., Shang, C.C., Cao, D.Y., Tian, T.T. & Zhu, X.Q. (2011). Seroprevalence of *Toxoplasma gondii* infection in dairy goats in Shaanxi Province, Northwestern China. *Parasites* & *Vectors* 4: 47.