

Antiviral activities of Cholistani plants against common poultry viruses

Shahzad, M.I.^{1*}, Anwar, S.², Ashraf, H.², Manzoor, A.², Naseer, M.², Rani, U.², Aslam, Z.², Saba, N.², Kamran, Z.³, Ali, S.⁴, Aslam, J.¹ and Arshad, M.⁵

¹Department of Biochemistry & Biotechnology. The Islamia University of Bahawalpur, Pakistan

²Department of Botany. The Govt Sadiq College Women University, Bahawalpur, Pakistan. Pakistan

³University college of Veterinary and Animal Sciences, The Islamia University, Bahawalpur

⁴Govt. poultry farm, Model Town A, Bahawalpur. Pakistan

⁵Department of Biochemistry, College of Veterinary and Animal Sciences, Jhang Campus, UVAS Lahore, Pakistan

*Corresponding author e-mail: mirza.inran@iub.edu.pk

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Abstract. Herbal medicines are becoming more popular and acceptable day by day due to their effectiveness, limited side effects, and cost-effectiveness. Cholistani plants are reported as a rich source of antibacterial, antifungal, antiprotozoal, antioxidant, and anticancer agents. The current study has evaluated antiviral potential of selected Cholistani plants. The whole plants were collected, ground and used in extract formation with *n*-hexane, ethyl acetate and *n*-butanol. All the extracts were concentrated by using a rotary evaporator and concentrate was finally dissolved in an appropriate vol of the same solvent. All of the extracts were tested for their antiviral potential by using 9-11 days old chick embryonated eggs. Each extract was tested against the Avian Influenza virus H9N2 strain (AIV), New Castle Disease virus Lasoota strain (NDV), Infectious bronchitis virus (IBV) and an Infectious bursal disease virus (IBDV). Hemagglutination test (HA) and Indirect Hemagglutination (IHA) tests were performed for different viruses. The overall order of the antiviral potential of Cholistani plants against viruses was NDV>IBV>IBDV>AIV. In terms of antiviral activity from extracts, the order of activity was *n*-butanol>ethyl acetate>*n*-hexane. The medicinal plants *Achyranthes aspera*, *Neuroda procumbens*, *Panicum antidotale*, *Ochthochloa compressa* and *Suaeda fruticosa* were very effective against all four poultry viruses through their extracts. The low IC₅₀ values of these extracts confirm the high antiviral potential against these viruses. It is worth to mention that *Achyranthes aspera* was found positive against IBDV through all its extracts which overcome the problem of unavailability of any known drug against IBDV. In short, the study proved that Cholistani plants are rich source of antiviral agent and their extracts can be used as good source of antiviral drugs both in crude and in purified form.

INTRODUCTION

For thousands of years, medicinal plants are in use to support human health. The medicinal constituents of these plants are not only playing important roles in traditional medicines but also modern medicines (Arunkumar & Muthuselva, 2009). The drugs obtained from plants are considered relatively safer, cheap, easy to use and with no or limited side effects. The use of herbal medicines is increasing all over the world

(Rasool *et al.*, 2015). The use of plant extracts in food, cosmetics, and pharmaceutical industries shows its public acceptance (Nostro *et al.*, 2000). The trend to use natural products and medicinal plants in the USA and Canada is exponentially growing (Roeder *et al.*, 2015). Several hundred natural products have been isolated for screening and identifying the antiviral activity, and some have been successfully approved for pre-clinical and clinical trials (Kitazato *et al.*, 2007). Cholistani plant *Oxystelma*

esculentum (Asclepiadaceae) is a twiner growing in waterlogged areas and used commonly as anti-ulcer and diuretic (Pandya & Anand, 2013). *Haloxylon silicornicum* (Amaranthaceae) has values of vegetation (Shafi *et al.*, 2002). Its extracts are also used in the treatment of diabetes and eye disorders. *Haloxylon recurvum* (Chenopodiaceae) is used externally against insect bites and also acknowledge a good source of fuel (Dagla & Shekhawat, 2005). *Achyranthes asper* (Amaranthaceae) is best known for its cardiovascular and spermicidal activities (Srivastava *et al.*, 2011). *Solanum surattense* (Solanaceae) commonly known as Indian nightshade and has been used for curing fever, cough, asthma, and diabetes (Munuswamy *et al.*, 2013). *Salsola bryosoma* (Chenopodiaceae) used as fodder, fuel and fire set. This plant is also used in Mauritania for laundering cloth. *Neurada procumbens* (Neuradaceae) is well known for its common uses (Chen *et al.*, 2004). *Panicum antidotale* (Graminae) known as bansighass and used to control soil erosion (Arshadullah *et al.*, 2009). *Sporo bolusicolados* (Poaceae) commonly known as sakhm, mainly used as forage (Gulzar & Khan, 2015). *Ochthocloa compressa* (Poaceae) is used as the source of fodder (Ahmed *et al.*, 2014). *Suaeda fruticosa* (Amaranthaceae) is shrubby halophyte and resist Reactive Oxygen Species (Samiullah *et al.*, 2012). Several plants are found effective against herpes simplex type I, Avian Influenza strain, and other poultry viruses (Abad *et al.*, 1997; Chollom *et al.*, 2012; Pant *et al.*, 2012; Chen *et al.*, 2014;). A highly effective natural product based antiviral compound against the hepatitis C virus is reported by (Lee *et al.*, 2016). Similarly, Cholistani plants are found effective against the Infectious Bronchitis Virus (IBV) and Infectious Bursal Disease Virus (IBDV) (Aslam *et al.*, 2016).

Poultry viruses are a big cause of economic loss to industry due to their high morbidity and mortality in birds. Viruses like New Castle Disease (NDV), Avian Influenza Virus (AIV) H9N2, Infectious Bronchitis Virus (IBV) and Infectious Bursal Disease Virus (IBDV) are a serious threat to the poultry industry (Aslam *et al.*, 2016). Newcastle

disease (ND), caused by virus of family Paramyxoviridae and is one of the most urgent and serious problem of poultry. The NDV LaSota strain is naturally occurring low-virulence strain, has been routinely used as a live vaccine throughout the world (Zhao *et al.*, 2016). Influenza A viruses are members of the *Orthomyxoviridae* family and contain a segmented, negative-sense RNA genome encoding 10 proteins and a variable number of accessory proteins. Influenza A viruses are commonly characterized by their combinations of surface proteins, haemagglutinin (HA) and neuraminidase (NA), giving rise to a multitude of different subtypes designated like H9N2 (Wu *et al.*, 2014). Infectious bronchitis virus (IBV), which cause one of the major and economically important disease of poultry is commonly known as IB or Muttadi Khansi. Interestingly, it was found that IBV is able to inhibit multiple cellular stress granule signaling pathways, at the same time, its replication also results in the induction of stress granules in certain parts of infected cells (Brownsword *et al.*, 2020). Infectious bursal disease (IBD) is commonly known as Gumboro disease is a very common disease of young chick, caused by IBD virus. Its infection causes loss of B cells approximately 7-10 days post infection. The socio-economic impact of IBDV is worldwide (Vanden Berg *et al.*, 2000).

Every year these viruses are causing great economic loss and affecting the trade of poultry all over the world (Okwor *et al.*, 2011; Aslam *et al.*, 2016; Sae Silva *et al.*, 2016). Extensive poultry farming, dense human population, lack of education, a high risk of infection and poor biosecurity measures are common factors in spread of viral diseases in Southeast Asian, North African and Middle East countries (Premnathan *et al.*, 1992; Giasuddin *et al.*, 2012; Lee *et al.*, 2012; Li *et al.*, 2016). Although extensive vaccination campaigns were introduced to control viral outbreaks but still these diseases are uncontrolled in different parts of the world especially in Pakistan. The phenomenon like genetic drifting/genetic shifting, availability of very few antiviral agents and poor quality of vaccines are other

factors to avoid control of these diseases (Singh, 2011). Therefore, the demand of new, novel and more effective antiviral agent(s) is increasing. The disease like IBD, there is no known drug is available in the market except crude extracts of medicinal plants (Gupta & Chaphalkar, 2015; Aslam *et al.*, 2016; Shahzad *et al.*, 2019).

MATERIALS AND METHODS

Collection of specimens

Eleven fresh plants including *Oxystelma esculentum* L.F. (Asclepiadaceae), *Haloxylon salicornicum* Moq. (Amaranthaceae), *Haloxylon recurvum* Moq. (Chenopodiaceae), *Achyranthes aspera* ITIS. (Amaranthaceae), *Solanum surattense* Linn. (Solanaceae), *Salsola baryosma* Roem. et Shult (Chenopodiaceae), *Neuroda procumbens* L. (Neuradaceae), *Panicum antidotale* Retz. (Poaceae), *Sporobolus icolados* Nees ex Trin. (Poaceae), *Ochthochloa compressa* CoL (Poaceae), and *Suaeda fruticose* auct. non Forsk (Amaranthaceae), were collected from the Cholistan desert and identified by Botanists at Herbarium of the Cholistan Institute of Desert Studies (CIDS), The Islamia University of Bahawalpur, as a whole plant. All the vouchers are available at CIDS, The Islamia University of Bahawalpur. Four poultry viruses were selected for this study, which includes vaccinal strains of New Castle disease virus (NDV) named as LaSota, Avian influenza virus (AIV) H9N2 named as Gallimune Flu H9 M.E from Merial Laboratories Italy. Infectious bronchitis virus (IBV) H120 from IZO S.U.R.L99/A-25124 Brassica Italy. Infectious bursal disease virus (IBDV) named as DS Gumboro Vac from Dae Sung Microbiological Laboratories, Seoul, South Korea.

Preparation of Plant extracts

Whole plants were collected, cleaned and dried under the shade at RT for 10-12 days. The dried plants were ground to powder form and powders were kept in airtight containers at RT. Ten gram powder from each plant was taken and dissolved in 100 mL *n*-hexane,

Ethyl acetate, and *n*-butanol. The methanol extracts of these plants were already tested by author (Aslam *et al.*, 2016; Shahzad *et al.*, 2019). The solutions were kept in airtight containers for 96h at constant shaking and subjected to the rotary evaporator (45-50°C). After evaporating the solvent the solutes were rinsed by methanol and chloroform in conical flasks and allow them to re-evaporate. Finally, the precipitates were weighed and dissolved in each corresponding solvent @ 0.5mg/mL. The extracts were sterilized through syringe filter and stored at -20°C (Joshi, 2013).

Viruses

The four viruses including Avian Influenza virus H9N2 strain (AIV), New Castle Disease virus Lasoota strain (NDV), Infectious bronchitis virus (IBV) and an Infectious bursal disease virus (IBDV) were used in this study. All the viruses were tested against different extracts through *In Ovo* antiviral assay.

Inoculation of viruses in chicken egg

Specific pathogen free (SPF) eggs were obtained from the government poultry farm, Model Town A. Bahawalpur. All the above-mentioned viruses were inoculated in the chorioallantoic fluid of 9-11 days old chick embryonated (CE) eggs. The eggs were candled before and after inoculations. The broader ends of eggs were swabbed with 70% alcohol and transferred into cleaned trays in biosafety cabinet II. The broader ends of eggs were drilled with sterile needle and viral inoculum is injected through sterile 3cc syringe. After inoculation, the hole was sealed with molten wax and eggs were incubated at 37°C. The eggs were harvested 72 h post-inoculation (PI) and allantoic fluids were collected and subjected to HA or IHA (in case of IBDV) test to check titer of each virus. This whole procedure was done in biosafety cabinet type II. Serial passages of viruses were done to increase titer before taking antiviral trials from extracts of Cholistan plants (Bajpai & Chandra, 1990).

Hemagglutination Test (HA)

The HA test was performed to quantify the titer of viruses except for IBDV (Elizondo-

Gonzalez *et al.*, 2012). A freshly prepared Alsevier solution was used to collect 5 mL of chicken blood. The blood was centrifuged at 4000 rpm for 5 min and supernatant was discarded. The cells were washed with PBS (pH 7.4) three times and finally dissolved to make a 1% solution. The 96 wells round bottom plates were used in assay. First of all, 50 μ L PBS solution was added in each well. Then 50 μ L virus (allantoic fluid) was added in first well and mixed three times and 50 μ L solution was transferred into 2nd well and mixed and this serial dilution was proceeded up to 11th well. The 12th well was kept as -ve control. The 1% RBC's solution was added @ 50 μ L in each well and plates were kept at 37°C for 1 hr and the HA activity was noted (OIE, 2012). In the case of AIV H9N2 and NDV the drug amantadine was used as positive control and in case of IBV LiCl used as positive control.

Indirect Hemagglutination (IHA) Test

This test was performed to quantify IBDV and human blood type "O" was used in it. Three mL blood was taken in a 4% sodium citrate solution and mixed gently. It was centrifuged at 4000 rpm for 5 min and the supernatant was removed. The cells were dissolved in 10 mL PBS (pH 7.4) and centrifuged again at 4000 rpm for 5 min. This step was repeated three times. After washing, the cells were sensitized by mixing with an equal vol of IBDV and two vol of PBS (pH 7.4). The contents were mixed gently and incubated at 37°C for 45 min. After incubation, the cells were centrifuged at 4000 rpm and the supernatant was removed and washed with PBS (pH 7.4) again and finally, diluted to make 1% suspension (Okwor *et al.*, 2011). Later, the standard HA test was performed by using these sensitized cells (Hussain & Gorski, 2004). No positive control used for IBDV because no known drug is available.

In Ovo Analysis

The filter sterilized plant extracts were mixed with equal vol. of viral inoculums and injected in 9-11 days old CEE (Rajbhandari, 2001). Autoclave distilled water was used in case of negative control and virus without any

plant extracts were used in case of virus control. All the inoculated eggs were harvested 72 h PI and subjected to HA or IHA. Any change in the case of button formation in comparison to virus control was noted. IC₅₀ of each positive extract was calculated by making serial dilutions of extracts and challenged against viruses.

Calculation of IC50

IC50 was calculated by testing serially diluted drugs against each virus. Easy fit software was used to calculate dose and time dependent curves by linear regression and IC50 value presented the results in Table 1.

Statistical Analysis

Data showed mean and standard deviations. Statistical analysis was performed using one-way ANOVA and a p-value considered a significant difference at P<0.05 (Steel *et al.*, 1997).

RESULTS

Cultivation of Viruses

Viruses were cultivated in embryonated eggs (CE) to achieve certain titer before the start of antiviral studies. In the case of New Castle Disease (NDV), the HA titer 2048 was achieved after the 6th passage. In the case of the Avian Influenza Virus (AIV) and Infectious Bronchitis Virus (IBV), the HA titers 2048 were achieved after the 9th passage. Similarly, in the case of Infectious Bursal Disease Virus (IBDV), the IHA titer 1024 was achieved after the 20th passage. Later these high titer viruses were subjected to *In Ovo* antiviral assays.

Anti-NDV activity from different extracts of Cholistani plants

According to this experiment, the *n*-hexane extracts of all plants were effective in controlling NDV. The *n*-hexane extracts of *A. aspera*, *S. baryosma*, *P. antidotale*, *H. salicornicum*, *O. esculentum*, *S. icolado*, *S. fruticosa*, *H. recurvum*, *N. procumbens* were completely and equally effective (HA titer 0) against NDV. The extract of *S.*

surattense was slightly less effective and it did not completely inhibit the growth of the virus (HA titer 16). The extract of *O. compresssa* was further less effective and it has reduced the HA titer at 64. According to the results of ethyl acetate extracts of Cholistani plants, all extracts were effective in controlling NDV. The ethyl acetate extract of *O. esculentum*, *S. icolados*, *A. aspera*, *S. baryosma*, *P. antidotale*, *S. surattense*, *N. procumbens*, *O. compresssa* were completely effective and did not allow the virus to grow at all (HA titer 0). The extracts of *H. salicornicum*, *S. fruticosa*, *H. recurvum*, were also effective but did not stop the complete growth of the virus (HA titer 8, 8 and 32 respectively). In the case of *n*-butanol extracts of Cholistani plants were effective, but in varying degree. The extracts of *A. aspera*, *S. baryosma*, *P. antidotale* were first in order and they have completely inhibited the growth of the virus (HA titer 0). The extracts of *O. esculentum*, *S. icolados*, *H. recurvum* were second in order and HA titer was 2 in their cases. The extracts of *H. salicornicum*, *S. surattense*, *N. procumbens*, *S. fruticosa*, *O. compresssa* were third in order and HA titers were 8,16,16,32 and 32 respectively. The overall trend was *n*-hexane extracts > ethyl acetate extracts > *n*-butanol

extracts (Fig. 1). IC₅₀ of each positive extract was calculated and reported in Table 1.

Anti-AIV activity form different extracts of Cholistani plants

The *n*-hexane extract of *P. antidotale* was completely effective (HA titer 0). The extracts of *H. recurvum*, *O. compresssa*, *N. procumbens*, *S. surattense* were less effective and HA titers from these plants were 32, 32, 32 and 512 respectively. The extracts of *A. aspera*, *O. esculentum*, *S. baryosma*, *S. fruticosa*, *H. salicornicum*, *S. icolados* were almost ineffective (HA titer 1024 minimum and 2048 maximum). According to the results of ethyl acetate extracts of Cholistani plants, all extracts were not equally effective in controlling AIV. The extract of *A. aspera* was highly effective in controlling the growth of AIV (HA titer 0). The extract of *O. esculentum*, *O. compresssa*, *S. baryosma*, *H. recurvum* were less effective (HA titer 64, 256, 256 and 512 respectively). The extract of *S. surattense*, *N. procumbens*, *H. salicornicum*, *P. antidotale*, *S. fruticosa*, *S. icolados* were not effective in controlling the growth of AIV. Similarly, in the case of *n*-butanol extract, all plants were not equally effective. The extract of *N. procumbens* was

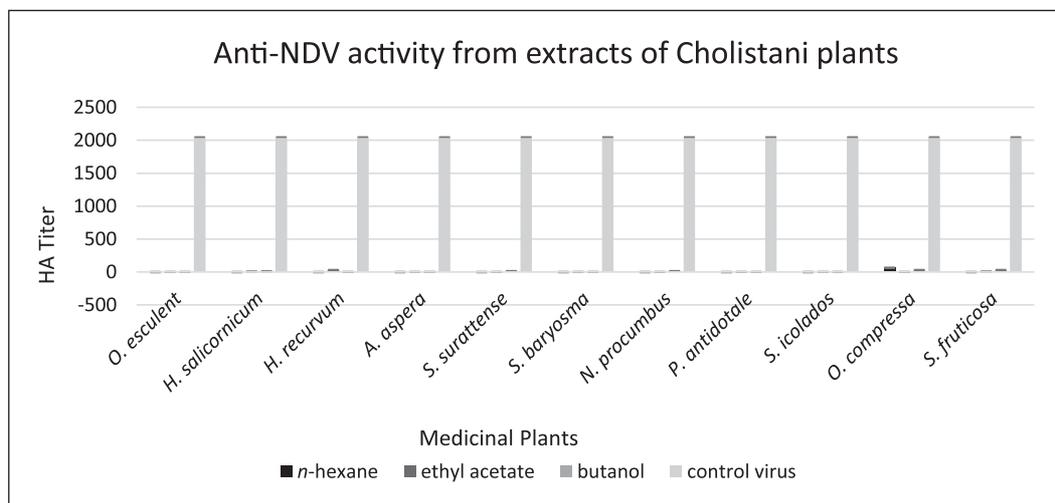


Figure 1. A: Anti-NDV activity from extracts of Cholistani plants.

1st bar of each column express the HA titer of *n*-hexane extract. 2nd bar of each column presents the HA titer against ethyl acetate extract. 3rd bar of each column shows the HA titer against *n*-butanol extract and 4th bar of each column represents the HA titer of control virus.

most effective and it has completely inhibited the growth of AIV (HA titer 0). The extracts of *S. surattense*, *O. compressa*, *S. fruticosa*, *S. baryosma*, *O. esculentum* were next in order and slightly less effective in controlling the growth of AIV (HA titer 8, 16, 16, 64 and 64 respectively). The extract of *H. salicornicum*, *A. aspera*, *P. antidotale*, *H. recurvum*, *S. icolados* were ineffective in controlling the growth of virus (HA titer 2048). The overall trend was *n*-butanol extracts >*n*-hexane extracts> ethyl acetate extracts in controlling AIV H9N2 growth (Fig. 2). IC₅₀ of each positive extract was calculated and reported in Table 1.

Anti-IBV activity form different extracts of Cholistani plants

According to this experiment, the *n*-hexane extract of *O. esculentum*, *N. procumbus*, *A. aspera*, *S. baryosma* were effective and completely active (HA titer 0) against IBV. The extract of *O. compressa*, *H. recurvum*, *P. antidotale*, *S. fruticosa*, *S. icolados*, *S. surattense* were less effective and their HA titers were 64 and 512 respectively. The extract of *H. salicornicum* was entirely ineffective (HA titer 2048). In the case of ethyl acetate extracts, some extracts were very

effective like extracts of *O. esculentum*, *N. procumbus*, *S. surattense*, *A. aspera*, *S. baryosma*, *H. salicornicum*, *S. icolados* (HA titer 0). The extract of *P. antidotale*, *S. fruticosa*, *O. compressa* were moderately effective (HA titer 8 and 32 respectively) and extract of *H. recurvum* was absolutely ineffective. In the case of *n*-butanol extracts all plants were effective, but in varying order. The extracts of *A. aspera*, *S. baryosma* were very effective (HA titer 0) but extracts of *O. compressa*, *O. esculentum*, *H. recurvum*, *S. surattense*, *S. icolados*, *P. antidotale*, *N. procumbus*, *S. fruticosa* were moderately effective and HA titers were 8, 64, 128, 128 and 256 respectively and extract of *H. salicornicum* was completely ineffective. The overall trend against this virus was ethyl acetate extracts > *n*-hexane extracts > *n*-butanol extracts (Fig. 3). IC₅₀ of each positive extract was calculated and reported in Table 1.

Anti-IBDV activity form different extracts of Cholistani plants

According to this experiment, the *n*-hexane extract of all plants was ineffective in controlling IBDV expect *A. aspera* (IHA titer 64), which was moderately active. In the

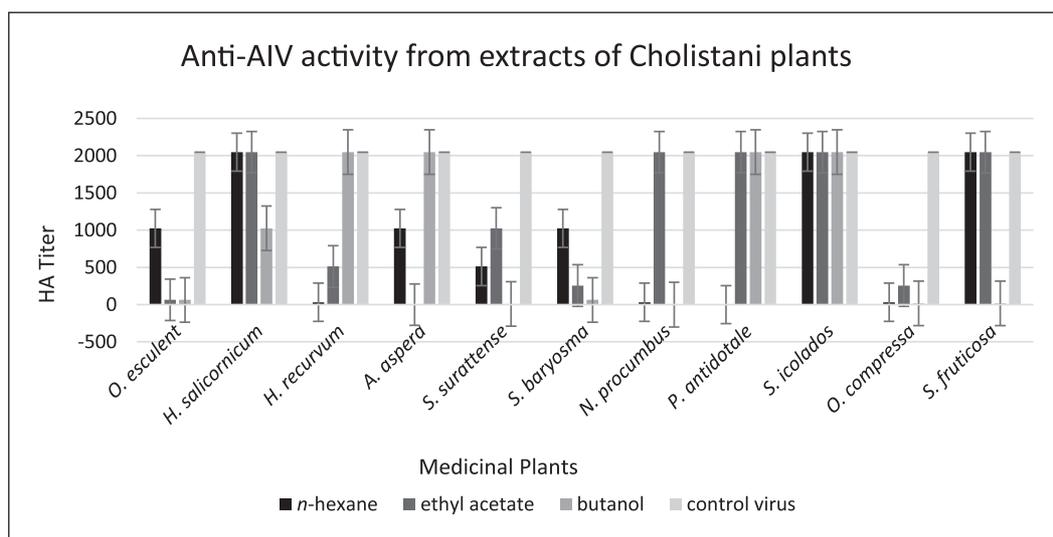


Figure 2. B: Anti-AIV activity from extracts of Cholistani plants.

1st bar of each column express the HA titer of *n*-hexane extract. 2nd bar of each column presents the HA titer against ethyl acetate extract. 3rd bar of each column shows the HA titer against *n*-butanol extract and 4th bar of each column represents the HA titer of control virus.

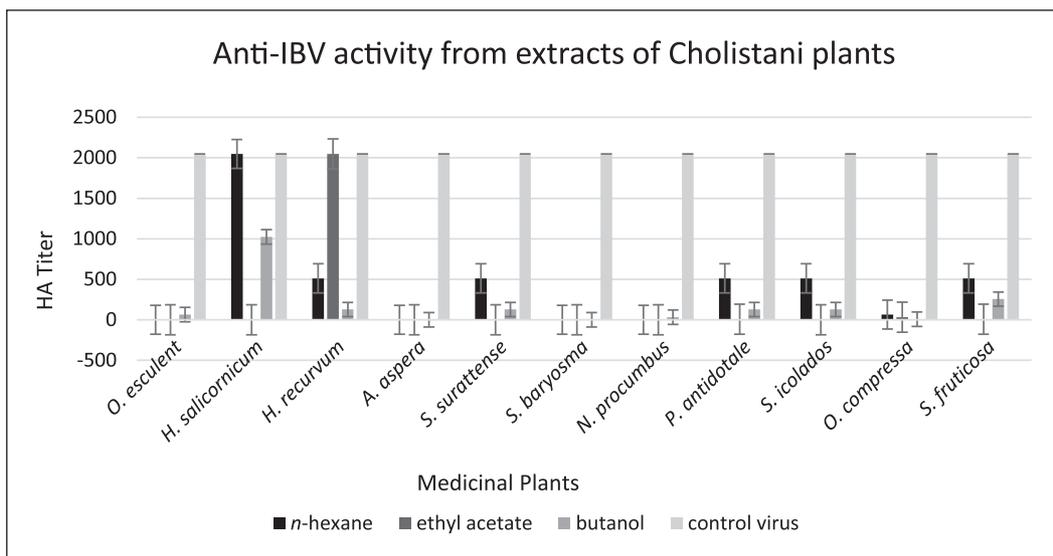


Figure 3. C: Anti-IBV activity from extracts of Cholistan plants. 1st bar of each column express the HA titer of *n*-hexane extract. 2nd bar of each column presents the HA titer against ethyl acetate extract. 3rd bar of each column shows the HA titer against butanol extract and 4th bar of each column represents the HA titer of control virus.

case of ethyl acetate extracts, some extracts were effective in controlling IBDV like *O. Compressa*, *N. procumbens* and some were moderately effective like *P. antidotale*, *H. recurvum*, *A. aspera* (IHA titer 16, 16, 64, 64 and 64 respectively) and others were finally ineffective. In the case of *n*-butanol extracts, *N. procumbens* was completely effective (IHA titer 0) and extracts of *O. Compressa*, *S. baryosma*, *P. antidotale*, *A. aspera*, *S. fruticosa* were moderately effective (IHA titers 8, 16, 32, 32 and 32 respectively) and the rest of plant extracts were totally ineffective. The overall trend against IBDV was *n*-butanol extracts > ethyl acetate extracts > *n*-hexane extracts (Fig. 4). IC₅₀ value of each extract was calculated and reported in Table 1.

DISCUSSION

According to the World Health Organization (WHO), more than 70% of populations of developing countries still rely on traditional medicines to resolve their health issues (Rabe & Van Staden, 2000). The use and importance of herbal medicines are on a

continuous rise for the last decade (Knezevic *et al.*, 2016). The use of traditional medicines and medicinal plants is embedded in the history of mankind. The extracts were originally done by crushing or squeezing or boiling herbal materials in water (Sujarwo, 2015). The plant-based products are more effective and work synergistically and provide less time for microbes to develop resistance against it (Solanki, 2010). Hundreds of plants and herbs have been reported with significantly high antimicrobial potentials. Some phytochemicals, including flavonoids, terpenoids, polyines, thiophenes, polyphenolics, lignans, sulphides, coumarins, saponins, furyl compounds, alkaloids, proteins, and peptides have been reported from different plants with antiviral potentials against different viruses (Jassim & Naji, 2003; Shahzad *et al.*, 2019). Most of these compounds are secondary metabolites of plants and they are not only effective against the virus but also bacteria and fungi (Gupta & Chaphalkar, 2015). The other compounds like volatile essential oils from spices and herbal teas are also reported with antiviral activities (Jassim & Naji, 2003). Typically 20–30% of plants from temperate regions have

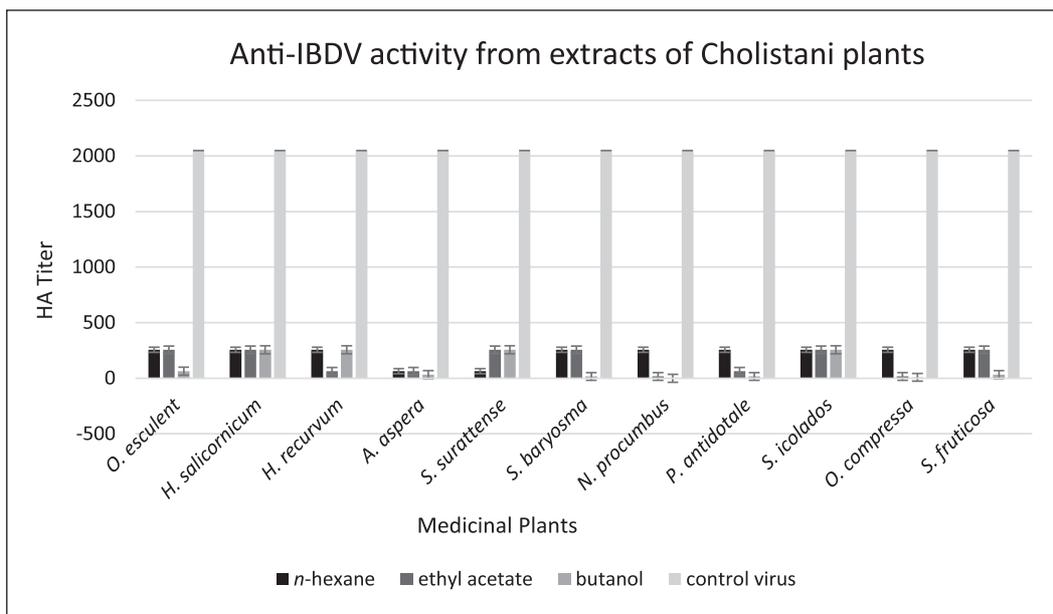


Figure 4. D: Anti-IBV activity from extracts of Cholistani plants. 1st bar of each column express the HA titer of *n*-hexane extract. 2nd bar of each column presents the HA titer against ethyl acetate extract. 3rd bar of each column shows the HA titer against *n*-butanol extract and 4th bar of each column represents the HA titer of control virus.

been observed to possess antiviral activity. A large number of compounds of varied chemical structures isolated from medicinal plants have been shown to possess antiviral activity (Perez, 2003).

According to this study no. of selected Cholistani plants were found effective against common poultry viruses. In the case of Newcastle disease virus (NDV), strong anti-NDV activities were reported from Cholistani plants. According to this study, 9 out of 11 *n*-hexane, 9 out of 11 ethyl acetate and 9 out of 11 *n*-butanol extracts were positive against NDV. Very low values of IC_{50} of plants like *A. aspera*, *S. baryosma*, *N. procumbus*, *P. antidotale*, and many others depict the potential of the Cholistani plant against NDV. The results of this study are in accordance with already reported studies like (Rajbhandari *et al.*, 2001) have reported antiviral activities from selected medicinal plants by *In Ovo* antiviral assays. (Elizondo-Gonzalez *et al.*, 2012) have reported a plant based compound Fucoidan from *C. Okamuraanus* against NDV. (Premnathan *et al.*, 1992) has reported antiviral activity

against NDV, vaccinia and HBV from methanol extracts of different seaweeds, sea grasses and of mangroves of the southeast coastal region of India. Similarly, in another study done by (Usha *et al.*, 2012) aqueous, ethanol and methanol extracts of *C. crista* L. were tested against NDV. (Chollom *et al.*, 2012) have reported anti-NDV activity from fruit pulp and leaf extracts of *Momordic abalsamin* through the cell line bases antiviral assay.

In the case of Avian Influenza Virus (AIV) H9N2, number of Cholistani plants were effective in different fractions. Total 4 out of 11 *n*-hexane, 2 out of 11 ethyl acetate and 6 out of 11 *n*-butanol extracts were effective. Only *O. compressa* was effective in controlling AIV in all of its fractions, the rest of the plants were effective in one or two fractions. Similarly, the low IC_{50} value of these plants confirms the potential of Cholistani plants against AIV. The similar results are reported by other researchers like (Park, 2003) have reported anti AIV activity from methanol extracts of four Korean medicinal plants. (Lee *et al.*, 2016; Shahzad

et al., 2019) have reported anti AIV activity from n-hexane and ethyl acetate extracts of green tea. Studies done by (Droebner *et al.*, 2017; Ehrhardt *et al.*, 2007) have confirmed the antiviral effects of *Proantho cyanidins* derived from plants against AIV strains.

According to the results of this study, Cholistani plants are rich sources of anti-IBV agents and 5 out of 11 *n*-hexane, 10 out of 11 ethyl acetate and 5 out of 11 *n*-butanol extracts were positive against IBV. Overall 5 plants including *O. esculent*, *A. aspera*, *S. baryosma*, *N. procumbus* and *O. compressa* were completely effective through all of their extracts in controlling IBV even very low IC₅₀ depict the potential of these plants against IBV. (Chen *et al.*, 2014) have reported antiviral activity against IBV from ethanol extracts of *R. rosea* roots, *N. sativa* seeds, and *S. snigra* fruit. (Ahmed *et al.*, 2014; Musaddiq *et al.*, 2020) have reported anti-IBV and IBDV activity from methanol extracts of Cholistani plants. In the case of Infectious Bursal Disease Virus (IBDV), 2 *n*-hexane, 5 ethyl acetate and, 7 *n*-butanol extracts were effective. Overall *A. aspera* was found very effective against IBDV through all of its extracts. The low IC₅₀ values of extracts of this plant have highlighted its potential against IBDV. It is worth mentioning that there is no known anti IBDV drug available in the market and this disease is known as AIDS of poultry. The number of studies are available, which have highlighted the use of medicinal plants against IBDV. (Pant *et al.*, 2012) have reported anti IBDV activity from the hydro-alcohol extract of *W. somnifera* roots. (Ahmed *et al.*, 2014) have reported anti IBDV activity from ethanol extracts of *M. oleifera*, *P. emblicus*, *G. glabra*, and *E. jambolana* through cell line based assay (Simoni *et al.*, 2007) have tested number of medicinal plants from Brazilian flora against IBDV, herpes virus type I (BoHV-1) and avian reovirus. A total 8 out of 16 plants were effective against (BoHV-1) and avian reovirus but none of them was effective against IBDV. Anti-IBV and IBDV activity from methanol extracts of Cholistani plants. The mechanism of most of the antiviral compounds is needed to be determined yet (Aslam *et al.*, 2016; Shahzad *et al.*, 2019). Some researchers

have anticipated the loss with the attachment of the virus to the host cell surface by antiviral agents, especially at the early stage of infection and some have suggested complete or partial inhibition of essential enzymes of viruses. Some other researchers have suggested the protein or polypeptide nature of antiviral compounds (Bajpai & Chandra, 1990).

Vanden *et al.* (1986) have reported that there will be an increasing need for substances with antiviral activity soon because the current treatments of viral infections are often unsatisfactory and limited. Mutation in viruses and resistance to the existing antiviral agents will readily arise upon treatment. These compounds could be extracted from higher plants. Many of these plants have been used historically to treat diseases now known to be of viral origin (Louis & Balakrishnan, 1996; Tahir *et al.*, 2019). The author has already published the effect of the methanol extract of these plants against the same poultry viruses (Aslam *et al.*, 2016; Shahzad *et al.*, 2019; Musaddiq *et al.*, 2020).

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Declaration of interest

All authors are duly informed and gone through this manuscript. There is no conflict of interest in this paper.

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